

## Letter to Editor

JMR 2016; 2(1): 01-02 January- February ISSN: 2395-7565 © 2016, All rights reserved www.medicinearticle.com

## Saliva in diagnostics- a useful screening tool

Jayita Poduval

Associate Professor, Department of ENT, Pondicherry Institute of Medical Sciences, Kalapet, Puducherry- 605014, India

The use of saliva for the purpose of diagnosing disease is scarcely known but is rapidly gaining popularity. In fact, the use of human saliva for diagnostics is changing from classical forms of testing such as that for salivary flow, to that of looking at salivary biomarkers. This article briefly reviews the same while differentiating between the two methods of diagnosis.

Salivary flow testing is also known as sialometry, which measures the flow rate of saliva. Saliva is mainly produced by the major salivary glands, viz. parotid, submandibular and sublingual glands. Salivary flow testing is usually included in the topodiagnostic methods of testing for facial nerve paresis or paralysis, and is a very sensitive early indicator of nerve damage because the flow is drastically reduced in the first 24 to 48 hours of nerve injury. This finding was earlier used to select patients for early facial nerve decompression but it is now well known that neuropraxia may cause such a temporary dysfunction of the nerve and does not reliably indicate the extent of the lesion. It has therefore been given up <sup>[1]</sup>, as have been the other topognostic tests, viz. Schirmer's test and electrogustometry.

The present indications of salivary flow testing are limited to the grading of severity of salivary gland disease, esp. Sjogren's Syndrome, and other similar autoimmune conditions, and in evaluation of their response to therapy. In these cases, it is usually combined with related tests like Schirmer's test and labial biopsy <sup>[2]</sup>. The co-operation of the patient is of utmost importance as the testing equipment needs to be installed accurately. Emotional factors may affect the test results, therefore, counselling and reassurance must be given to the patient along with an informed, written consent.

Partial surgical removal of the parotid gland decreases flow in the gland but is not associated with complaints of xerostomia. It may also cause a compensatory increase in secretion from other glands. Cunning et al reported a decrease in unstimulated salivary flow in patients undergoing unilateral submandibular gland excision with increased subjective complaints of xerostomia <sup>[3]</sup>. Unstimulated saliva is collected 2 hours after eating or after an overnight fast. Whole saliva (a total product of all salivary glands) can be measured by a variety of volumetric and gravimetric techniques, including drooling, spitting, suction and swab <sup>[4]</sup>. Isolated parotid secretions can be obtained by placement of a suction cup over the Stensen's duct, usually a Carlson-Crittenden cup <sup>[5]</sup> or Lashley cup.

Human saliva can also be tested for levels of steroid hormones such as cortisol. This is useful in diseases of the adrenal glands, such as Cushing's syndrome and Addison's disease, especially screening for the latter <sup>[6, 7]</sup>. It may be used by dermatologists when confronted with hyperpigmented skin lesions, which among other things, could be due to Addison's disease. Saliva cortisol levels may also be used for the evaluation of chronic stress whereas salivary alpha amylase could be increased in acute stress <sup>[8]</sup>. Other hormonesboth male and female- in the saliva, may be evaluated in conditions such as polycystic ovary syndrome, anovulation, menopause and hypogonadism <sup>[9]</sup>. Similarly, insulin resistance, metabolic syndrome and diabetes mellitus could also be evaluated using salivary biomarkers such as chromogranin A and salivary lysozyme <sup>[10, 11]</sup>. The expression of nitric oxide in saliva correlates well with levels of it in the blood and is thus useful for the detection and measurement of this biomarker that could be used for monitoring blood pressure and heart health <sup>[12, 13]</sup>.

The HIV antibody is also expressed in saliva and detection of the same is now approved for medical and commercial use <sup>[14]</sup>. Similarly, antibodies against Hepatitis A, B and C are also well evaluated using saliva <sup>[15, 16]</sup>. Periodontal disease is also another area where salivary biomarkers have proved useful.

Allergic states, parasitic and fungal infections can also be adequately assessed using saliva as a screening and diagnostic tool <sup>[17]</sup>. The same applies to the detection of illegal drugs as well as prescription medications, and the monitoring of circadian rhythms in certain industries and occupations. Breast, pancreatic and oral cancers could also be screened using saliva <sup>[18]</sup>.

\*Corresponding author: Dr. Jayita Poduval Associate Professor, Department of ENT, Pondicherry Institute of Medical Sciences, Kalapet, Puducherry-605014, India

## The Journal of Medical Research

PCR (polymerase chain reaction), ELISA (enzyme- linked immunosorbent assay), HRMS (high resolution mass spectrometry) are the more popular methods in current use when working with a saliva sample. Nanotechnology is believed to further enhance the sensitivity of these methods. Even though variability may exist among laboratories using saliva for diagnostics, the safety and non invasive nature, affordability and ease of collection allow fairly good levels of accuracy. Standardization is possible pending further research and widespread adoption of this modality of diagnostics.

## References

- 1. May M, Hardin WB Jr, Sullivan J. Natural History of Bell's Palsy:the Salivary Flow Test and other prognostic indicators. Laryngoscope 1976;86:704-12.
- 2. Scott-Brown's, Otolaryngology-Head and Neck Surgery:6th Edn, Vol 5 chapter-Non-neoplastic diseases of salivary glands.
- Cunning DM, Lipke N, Wax MK. Significance of unilateral submandibular gland excision on salivary flow in noncancer patients. Laryngoscope 1998;108(6):812-5.
- 4. Robert Lee Witt. In Salivary Gland Diseases: Surgical and Medical Management, Thieme 2005, p 35.
- Shannon IL, Prigmore JR, Chauncey HH. Modified Carlson-Crittenden Device for the collection of Parotid Fluid. JDR(Journal of Dental Research) 1962; 778-83.
- Sakihara S, Kageyama K, Oki Y, Doi M, Iwasaki Y, Takayasu S et al. Evaluation of plasma, salivary, and urinary cortisol levels for diagnosis of Cushing's syndrome. Endocr J 2010;57(4):331-7.
- Restituto P, Galofré JC, Gil MJ, Mugueta C, Santos S, Monreal JI et al. Advantage of salivary cortisol measurements in the diagnosis of glucocorticoid related disorders. Clin Biochem 2008 Jun;41(9):688-92.
- Takai N, Yamaguchi M, Aragaki T, Eto K, Uchihashi K, Nishikawa Y. Effect of psychological stress on the salivary cortisol and amylase levels in healthy young adults. Archives of Oral Biology 2004;49: 963–8.

- Gavrilova N, Lindau ST. Salivary sex hormone measurement in a national, population-based study of older adults. J Gerontol B Psychol Sci Soc Sci 2009 Nov;64 Suppl 1:i94-105.
- Soell M, Feki A, Hannig M, Sano H, Pinget M, Selimovic D. Chromogranin A detection in saliva of type 2 diabetes patients. Bosn J Basic Med Sci. 2010 Feb;10(1):2-8.
- Qvarnstrom M, Janket SJ, Jones JA, Jethwani K, Nuutinen P, Garcia RI, Baird AE, Van Dyke TE, Meurman JH. Association of salivary lysozyme and C-reactive protein with metabolic syndrome. J Clin Periodontol. 2010 Sep;37(9):805-11.
- 12. Hezel MP, Weitzberg E. The oral microbiome and nitric oxide homoeostasis. Oral Dis. 2015 Jan;21(1):7-16.
- Machha A, Schechter AN. Dietary nitrite and nitrate: a review of potential mechanisms of cardiovascular benefits. Eur J Nutr. 2011 Aug;50(5):293-303.
- Pascoe SJ, Langhaug LF, Mudzori J, Burke E, Hayes R, Cowan FM. Field evaluation of diagnostic accuracy of an oral fluid rapid test for HIV, tested at point-of-service sites in rural Zimbabwe. AIDS Patient Care STDS 2009 Jul;23(7):571-6.
- Tourinho RS, Amado LA, Villar LM, Sampaio DV, Moraes AC, Rodrigues do Ó KM, Gaspar AM, de Paula VS. Importance of the cutoff ratio for detecting antibodies against hepatitis A virus in oral fluids by enzyme immunoassay. J Virol Methods 2011 May;173(2):169-74.
- Yaari A, Tovbin D, Zlotnick M, Mostoslavsky M, Shemer-Avni Y, Hanuka N, Burbea Z, Katzir Z, Storch S, Margalith M. Detection of HCV salivary antibodies by a simple and rapid test. J Virol Methods 2006 Apr;133(1):1-5.
- Böttcher MF, Häggström P, Björkstén B, Jenmalm MC. Total and allergenspecific immunoglobulin A levels in saliva in relation to the development of allergy in infants up to 2 years of age. Clin Exp Allergy 2002 Sep;32(9):1293-8.
- Pink R, Simek J, Vondrakova J, Faber E, Michl P, Pazdera J, Indrak K. Saliva as a diagnostic medium. Biomed Pap Med Fac Univ Palacky Olomouc Czech Repub 2009 Jun;153(2):103-10.