

# **Research Article**

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# Comparative Study of the Phytochemical and Bio-activities of the Essential Oils from Ripe and Unripe Seeds of *Azadirachta indica*

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# Abstract

This study determines the secondary metabolites of the essential oils of ripe and unripe seeds of *Azadirachta indica* and then evaluated their antioxidant and antimicrobial potentials. Ripe and unripe seeds were subjected to hydrodistillation using a Clevenger-type apparatus and analyzed using gas chromatography and gas chromatography-mass spectrometry (GC-MS). Antioxidant and antibacterial activities of the volatile oils were also investigated using 2,2diphenyl-1-picrylhydrazyl (DPPH) and agar well diffusion methods, respectively. The GC-MS analysis showed that the essential oils of ripe and unripe seeds contained fourteen (14) and twenty-three (23) therapeutically active compounds, respectively. Compounds present in high quantity in the essential oil of ripe seeds were: 5-hydroxymethyltetrahydro-2furanol (35.5%) and 2,5-dimethyl-1,5-heptadiene-3,4-diol (11.8%), palmitic acid (5.0%) and methyl-9-octadecenoate (5.0%), while 2-methyl-2-pentanethiol (31.9%), *cis*-oleic acid (21.0%), 4-methyl-5-nonanone (10.5%), toluene (6.0%) and *o*-xylene (6.0%) were the principal compounds in the essential oil of the unripe seeds. Essential oils of both ripe and unripe seeds showed high inhibition against *Staphylococcus aureus*. The essential oil of the unripe seeds showed moderate to high inhibition against *Pseudomonas aeruginosa*. Free radical scavenging of the two essential oils gave IC<sub>50</sub> values of 2.00 and 2.50 for ripe and unripe seeds essential oil, respectively. Essential oil of unripe seeds has higher antimicrobial strength than that of the ripe seed. Essential oils of the seeds of *A. indica* could serve as a good source of pharmaceuticals and industrially useful compounds.

Keywords: Azadirachta indica, GC-MS, Phytochemical, Seeds essential oils, Antioxidant, Antibacterial.

#### INTRODUCTION

Plant essential oils and their compositions have multiple and varied therapeutic properties. They have received much attention due to their antioxidant potential in the prevention of reactive oxygen species (ROS) diseases [1-4]. Essential oils have special niche and great prospects as preservative and drug in food, nutraceutical and pharmaceutical industries. They are commonly used in complementary and alternative medicine [5,6].

Azadirachta indica is a tropical evergreen tree with frond-like leaves. It belongs to the family *Meliaceae*. It is a fast-growing tree with a height of 20–23 m, the trunk is straight and has a diameter around 4-5 ft. Its fruits are green drupes which turn golden yellow on ripening [7,8]. The plant contains secondary metabolites with immunomodulatory property; it is mainly used locally to reduce blood sugar levels and to treat various diseases such as malaria, cough, asthma, diabetics, rheumatism, leprosy, eye disorders, intestinal disorder, ulcers, urinary disorders, leprosy, hemorrhoids, cardiovascular diseases, gingivitis, kidney and livers problems, among many other medicinal uses [9-12]. Azadirachtin, a main component of most agrochemical is the most prominent constituent of the medicinal plant. Components of *A. indica* are used to regulate the metamorphosis and growth in insect from the larva to pupa stages. They are repellent, anti-feedant and offensive agent and induces sterility in insects by preventing oviposition and interrupting process of reproduction in insects [11, 13, 14].

To the best of our knowledge, there is no enough scientific information on the biological activities (free radical scavenging, antioxidant and antimicrobial potential) of the ripe and unripe seeds of this plant so far. Therefore, this research was undertaken with the aim of looking into the quantitative and qualitative properties of the essential oils of ripe and unripe seeds of *A*. *indica* from Nigeria.

#### **MATERIALS And METHODS**

# **Plant Materials**

Fresh ripe and unripe seeds of the plant were collected from Saint Mary Grammar School, Iwo, Osun state, Nigeria. The plant was authenticated as *Azadirachta indica* Linn at the Herbarium Department of the Forest Research Institute of Nigeria (FRIN), Ibadan.

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### **Extraction of Essential Oil**

Air-dried and pulverized (200 g) seeds of *A. indica* were subjected to hydrodistillation for 3 hrs using a Clevenger-type apparatus in accordance to British Pharmacopoeia methods [15]. The essential oils were dried using anhydrous sodium sulphate (Na<sub>2</sub>SO<sub>4</sub>), filtered and kept in vial and placed refrigerator regulated to 4  $^{\circ}$ C.

# **GC-MS** Analysis

GC-MS analysis of the essential oils was performed using a Shimadzu gas chromatograph model GCMS-QP2010 Plus (Japan) gas chromatographic (GC) system, equipped with a Mass selective detector and auto injector. Compounds were separated on capillary column RTx5ms-30 m x 0.25 mm x 0.25 µm film (5% diphenyl-95% dimethylpolysiloxane). A sample of 1.0  $\mu$ l was injected using the split mode (split ratio 1:100). For GC/MS detection, an electron ionization system, with ionization energy of 70 eV, was used. Column oven temperature was programmed from 80-220 °C at the rate of 4 °C min<sup>-1</sup>; initial and final temperatures were held for 3 and 10 minutes, respectively. Helium was used as a carrier gas at a flow rate of 1.5 ml min-1. Mass scanning range was m/z 40-700 while injector and MS transfer line temperatures were set at 220 °C and 290 °C, respectively. Quantification was completed by built-in data-handling software supplied by the manufacturer of the gas chromatograph. The results (composition) were reported as a relative percentage of the total peak area. Identification of the individual components was made by matching their recorded mass spectra with the NIST library stored in the computer which is dedicated to the GC-MS, the retention indices of the components were also compared with those of authentic compounds or with literature.

#### Antioxidant and Free Radical Scavenging Determination

Essential oils obtained from ripe and unripe seeds of *A. indica* were evaluated for antioxidant activity using DPPH. 1.0 mL of each of the essential oils solution and that of the control, ascorbic acid at different concentrations (1000, 100 and 10  $\mu$ gmL<sup>-1</sup>) in methanol were added to 1.0 mL of a 0.004% w/v methanol solution of DPPH and allowed to react at room temperature for 30 min. DPPH in methanol (2.5 mL) was used as a blank and ascorbic acid served as positive control. The absorbance of each solution was measured at 517 nm.

The percentage radical inhibition was evaluated based on the following expression:

$$I\%_{DPPH} = \frac{A_{blank} - A_{eo}}{A_{blank}} X \ 100$$

Where:  $A_{blank}$  and  $A_{eo}$  are the absorbance value for the blank and essential oil solutions, respectively. The dose-response curve was plotted and IC<sub>50</sub> value for the essential oils solutions and the standard were calculated [16].

#### **Screening of Antimicrobial Properties**

The essential oils of ripe and unripe seeds of *A. indica* were subjected to antimicrobial assays using agar well diffusion technique. *Staphylococcus aureus* (Gram-positive) and *Pseudomonas aeruginosa* (Gram-negative) were the bacteria strains used [17]. Nitrofuranton (NFT), a synthetic antibiotic was used as control. Sub-cultured bacterial isolates in nutrient broth was left for 18-24 hr to prepare bacteria suspension. The bacteria suspension (0.1 mL) prepared was inoculated into molten Mueller-Hinton agar medium at 45 °C and then poured

into sterile petri dish, the plate was allowed to set and wells were then bored into the agar medium using a sterile 6 mm cork borer. 10  $\mu$ L of the different concentrations of (1000, 100, and 10  $\mu$ gm L $^{-1}$ ) of each of the seeds essential oils and the control was added to each well. The plates were allowed to stand in the refrigerator for 1 hr to allow proper diffusion of the essential oil solution into the medium and then incubated at 37 °C for 18-24 hr after which they were observed for zones of inhibition [17, 18].

# **RESULTS AND DISCUSSION**

# Chemical Composition of Essential Oils of Ripe and Unripe Seeds of *A. indica*

The GC-MS analysis showed that the essential oils of ripe and unripe seeds of A. indica contained 14 and 23 components respectively. Compounds present in high quantity in the essential oil of ripe fruit were: 5-hydroxymethyltetrahydro-2-furanol (35.5%) and 2,5-dimethyl-1,5-heptadiene-3,4-diol (11.8%), palmitic acid (5.0%) and methyl-9octadecenoate (5.0%) while 2-methyl-2-pentanethiol (31.9%), cis-oleic acid (21.0%), 4-methyl-5-nonanone (10.5%), toluene (6.0%) and oxylene (6.0%) were the main component of the essential oil of the unripe seeds. Comparatively, both essential oils contained cis-oleic acid (ripe: 30.0%, unripe: 21.0%), palmitic acid (ripe: 5.0%, unripe; 3.0%), glyceryl-1,3-distearate (ripe: 3.7%, unripe; 2.0%), 1,2 dimethylbenzene (ripe: 1.7%, unripe: 6.0%), n-pentadecane (ripe:1.0%, unripe;1.0%), n-hexadecane (ripe:1.0%, unripe;1.0%), n-dodecane (ripe:1.0%, unripe;1.0%) and n-undecane (ripe:1.0%, unripe;1.0%). Important difference between the two essential oils was that sulphur containing compounds were present only in the essential oils of unripe seeds while 5-hydroxylmethyltetrahydro-2-furanol and 2,5-dimethyl-1,5heptadiene-3,4-diol were present only in the essential oil of ripe seeds of the plant. The chemical compositions of the essential oils obtained from the seeds investigated in this study were observed to be different from those obtained from other parts of the plant as reported by others scientists. The constituents of the leaf essential oil of A. indica from Egypt were mainly hydrocarbons (85.36%) and oxygenated compounds, mainly sesquiterpene oxide (5.04%). The constituents of the essential oil of flowers also included mainly hydrocarbons (63.22%) and oxygenated compounds which were unsaturated alcohols e.g farnesol (28.3%) [19]. According to Kamte et al. [17] and Dastan et al. <sup>[20]</sup>, the constituents of essential oil obtained from leaves of A. indica in the south of Iran were found to be different from that of the seeds analysed in this study.  $\gamma$ -elemene (20.8%), germacrene-B (20.3%), trans-carophyllene (13.5%), hexadecanal (12.8%) and methyl linoleate (10.5%) were the major compounds presents in the essential oil of the leaves. Okhale et al., (2018) [20] also reported that the main constituents of the essential oil obtained from roots of A. indica in Dikko, Niger State, Nigeria were: citronellic acid (29.60%), 1bromotriacontane (8.59%), totara-8,11,13-triene-7-6-13-diol (8.26%) and 4,8,12,15,15-pentamethyl-bicyclo[9.3.1]pentadeca-3,7-dien-12-ol (5.07%), these were also different from that of the seeds essential oils analyzed in this study. Oleic acid which is one of the principal component of the essential oils have been known to be an important dietary compound because it plays beneficial roles in human health; it improves heart conditions by lowering cholesterol and reducing inflammation [22], it also increases burning of fat which helps with weight loss [23], protects cells from free radical damage, prevent type 2 diabetes, prevents ulcerative colitis [24], generates brain myelin [25], involves in proper brain function [26, 27] and restores proper metabolism in failing hearts [28].

Compound	Retention Index	% Composition	
		Ripe EO	Unripe EO
2-methyl-2-pentenal	791	-	1.5
toluene	794	-	6.0
2-methyl-2-pentanethiol	837	-	31.9
β,β,β-trichloro- <i>tert</i> -butylalcohol	898	-	2.0
<i>o</i> -xylene	907	1.7	6.0
5-hydroxymethyltetrahydro-2-furanol	953	35.5	-
2-methyldecane	1051	1.0	-
4-methyl-5-nonanone	1087	-	10.5
n-undecane	1115	1.0	1.0
5-methylundecane	1150	-	1.0
2,2-dimethyl-1,5-heptadiene-3,4-diol	1193	11.8	-
n-dodecane	1214	1.0	1.0
n-tridecane	1313	1.0	-
3,5-diethyl-1,2,4-trithiolane	1344	-	1.0
n-pentadecane	1512	1.0	1.0
n-hexadecane	1612	1.0	1.0
1-butylhexylbenzene	1624	-	1.0
1-propyloctylbenzene	1643	-	1.0
n-heptadecane	1711	1.0	-
1-buthylnonylbenzene	1724	-	1.0
1-butyloctylbenzene	1731	-	1.0
1-propylnonylbenzene	1823	-	1.0
1-pentyloctylbenzene	1922	-	1.0
1-methyldecylbenzene	1933	-	1.0
palmitic acid	1968	5.0	3.0
methyl-9-octadecenoate	2102	5.0	-
cis-oleic acid	2175	30.0	21.0
methyl-10-octadecenoate	2085	-	2.0
glyceryl-1,3-distearate	4395	3.7	2.0
Percentage Total		99.7	98.9

# **DPPH Free Radical Scavenging and Antioxidant Properties**

The ability of the essential oils from the seeds of *A. indica* to scavenge free radical was determined on the basis of their concentrations [18], with  $IC_{50}$  values of 2.0 and  $2.5\mu gmL^{-1}$  for ripe and unripe seeds essential oils of *A. indica*, respectively and  $8.0\mu gmL^{-1}$  for ascorbic acid (the control). DPPH radical scavenging capability of the unripe seeds essential oil was a little lower than that of the ripe seeds essential oil. When hydrogen atom or electron was transferred to the odd electron in DPPH radical, the absorbance decreases proportionally to the increase of non-radical form. Lower absorbance of the reaction mixture (and thus lower  $IC_{50}$ ) indicates higher free radical scavenging activity. The antioxidant activities results of the two samples along with the positive control are shown in figure 1. Ability of the seeds to scavenge

DPPH radical that was determined showed that DPPH radical scavenging capability of the unripe seeds essential oil was a little lower than that of the ripe seeds essential oil. So it is possible that the other compounds in the unripe seeds essential oil also contributed to its reduced antiradical activity or that the presence of 5-(hydroxylmethyl)tetrahydro-2-furanol and 2,5-dimethyl-1,5-heptadiene-3,4-diol (which can donate hydrogen atom to the DPPH radical) account for higher antiradical activity of essential oil from the ripe seeds. This result was in agreement with the work of Alzohairy <sup>[10]</sup>, El-Hawary *et al.* [19], Elaigwu *et al.* <sup>[29]</sup> and Hossain *et al.* <sup>[30]</sup> which showed that the DPPH antioxidant of the essential oils of the leaves and flowers of *A. indica* were promising and were in relation with the chemical composition of the essential oils.

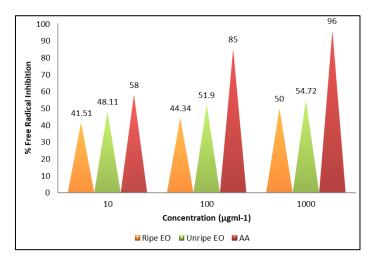


Figure 1: Antioxidant Properties of the Ripe and Unripe Seeds Essential Oils of A. indica

#### **Antibacterial Properties**

The antimicrobial activities of the essential oils of the ripe and unripe seeds of *A. indica* compared with the synthetic antibiotic against *S. aureus* and *P. aeruginosa* are shown in figure 2 and 3 below. The observed result showed that the essential oils from the ripe and unripe seeds of *A. indica* have similar antimicrobial activity against *S. aureus* and have higher activity than the synthetic antibiotic used (nitrofuraton). The essential oil of ripe seeds showed no antibacterial action against *P. aeruginosa* but that of the unripe seeds showed moderate to high antimicrobial activities of the essential oils of the seeds investigated in this study have higher antibacterial potential

compared to the antibacterial activity of ethanolic leaf extract of *A. excelsa* which exhibited weak inhibitory effect on *Shigella sonnei* (7.8-11.8 mm) and no inhibitory effect was observed against *Escherichia coli* and *Salmonella typhirium*. The investigated essential oils showed strong activities against multi- drug resistant bacteria due to the phytochemicals in the essential oil as well as the possible synergistic interaction between phytochemicals to penetrate the cell membrane of the organisms, inhibit their growth and proliferation; and inducing toxic effects to the membrane structures. The investigated essential oil as a natural antibiotic substance is locally available, easily accessible, easy to extract, inexpensive, environmentally safe and friendly [31, 32].

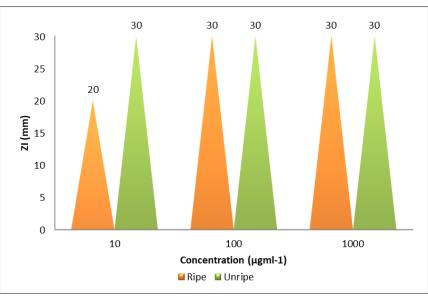


Figure 2: ZI (mm) of ripe and unripe Seeds essential oils of A. indica against S. aureus

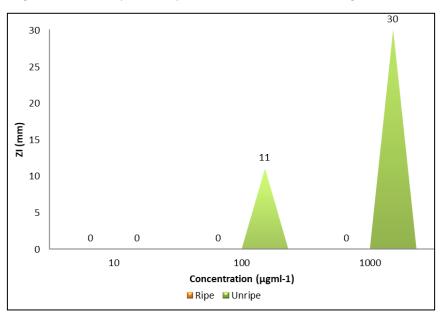


Figure 3: ZI (mm) of ripe and unripe Seeds essential oils of A. indica against P. aeruginosa

# CONCLUSION

The essential oils obtained from the ripe and unripe seeds of *A. indica* possessed significant antiradical and antimicrobial capacities indicating that essential oils from the seeds had multiple activities such as free radical scavenging, antioxidant and antimicrobial properties. Therefore the essential oils can be used for therapy against various oxidative stress diseases. The seeds from the plant can also be used as a source of oral drugs to fight infections caused by susceptible bacteria. Hence, essential oils from the seeds of *A. indica* should be treated as potential

natural free radical scavengers that may be of great use to the development of novel drugs and preservatives in food, nutraceutical and pharmaceutical industries.

#### **Conflict of interest**

We declare no conflict of interest.

#### REFERENCES

- Wang J, Liu H, Zhao J, Gao H, Zhou L, Liu Z, Chen Y and Sui P. Antimicrobial and Antioxidant Activities of the Root Bark Essential Oil of Periplocasepiumand Its Main Component 2-Hydroxy-4methoxybenzaldehyde. Molecules. 2010; 15:5807-5817.
- Ololade ZS, Fakankun OA, Alao FO and Udi OU. Phytochemical and Therapeutic Studies of the Fruit Essential Oil of Thuja orientalis from Nigeria, Global Journal of Science Frontier Research. 2014; 14(7):15-20.
- Chouhan S, Sharma K and Guleria S. (2017). Antimicrobial Activity of Some Essential Oils-Present Status and Future Perspectives. Medicines (Basel, Switzerland). 2017; 4(3):1-21.
- Sharifi-Rad J, Sureda A, Tenore GC, Daglia M, Sharifi-Rad M, Valussi M, Tundis R, Sharifi-Rad M, Loizzo MR, Ademiluyi AO, Sharifi-Rad R, Ayatollahi SA and Iriti M. Biological Activities of Essential Oils: From Plant Chemoecology to Traditional Healing Systems. Molecules (Basel, Switzerland). 2017; 22(1):1-55.
- Atanasov AG, Waltenberger B, Pferschy-Wenzig EM, Linder T, Wawrosch C, Uhrin P, Temml V, Wang L, Schwaiger S, Heiss EH, Rollinger JM, Schuster D, Breuss JM, Bochkov V, Mihovilovic MD, Kopp B, Bauer R, Dirsch VM and Stuppner H. Discovery and resupply of pharmacologically active plant-derived natural products: A review. Biotechnology advances. 2015; 33(8):1582-1614.
- Yuan H, Ma Q, Ye L and Piao G. The Traditional Medicine and Modern Medicine from Natural Products. Molecules (Basel, Switzerland). 2016; 21(5):1-18.
- Ogbuewu IP, Odoemenam VU, Obikaonu HO, Opara MN, Emenalom OO, Uchegbu MC, Okoli IC, Esonu BO and Iloeje MU. The Growing Importance of Neem (Azadirachta indica A. Juss) in Agriculture, Industry, Medicine and Environment: A Review. Research Journal of Medicinal Plants. 2011; 5:230-245.
- 8. Kumawat KR and Kumar R. Pharmacological and Therapeutical overview of neem (Azadirachta indica): A nature's drugstore, International Journal of Chemical Science. 2018; 2(2):23-27.
- Paul R, Prasad M and Sah NK. Anticancer biology of Azadirachtaindica L (neem): A mini review. Cancer Biology and Therapy. 2011; 12:467-476.
- 10. Alzohairy MA. Therapeutics Role of Azadirachta indica (Neem) and Their Active Constituents in Diseases Prevention and Treatment. Evidence-based complementary and alternative medicine. 2016; 16:1-11.
- Chaudhary S, Kanwar RK, Sehgal A, Cahill DM, Barrow CJ, Sehgal R and Kanwar JR Progress on Azadirachta indica Based Biopesticides in Replacing Synthetic Toxic Pesticides. Frontiers in Plant Science. 2017, 8
- Gupta A, Verma UP, Lal N and Ojha SK. Evolution and Exploration of Azadirachta indica in Dentistry: An Update, British Journal of Medicine and Medical Research. 2017; 21(8):1-15.
- 13. Mordue AJ and Nisbet AJ. Azadirachtin from the Neem Tree Azadirachta indica: its Action Against Insects. Anais da Sociedade Entomologica do Brasil, 2000; 29(4):615-632.
- 14. Hikal WM, Baeshen RS and Ahl HAHS. Botanical insecticide as simple extractives for pest control, Cogent Biology. 2017; 3:1-16.
- 15. British Pharmacopoeia British Pharmacopoeia Commission Publisher Stationery Office Books, 1993.

- Olugbami JO, Gbadegesin MA, and Odunola OA. In vitro free radical scavenging and antioxidant properties of ethanol extract of Terminalia glaucescens. Pharmacognosy research. 2015; 7(1):49– 56.
- 17. Kamte S, Ranjbarian F, Campagnaro GD, Nya P, Mbuntcha H, Woguem V, Womeni HM, Ta LA, Giordani C, Barboni L, Benelli G, Cappellacci L, Hofer A, Petrelli R and Maggi, F. Trypanosoma brucei Inhibition by Essential Oils from Medicinal and Aromatic Plants Traditionally Used in Cameroon (Azadirachta indica, Aframomum melegueta, Aframomum daniellii, Clausena anisata, Dichrostachys cinerea and Echinops giganteus). International Journal of Environmental Research and Public Health, 2017; 14(7):1-16.
- Njimoh DL, Assob JC, Mokake SE, Nyhalah DJ, Yinda CK and Sandjon B. Antimicrobial Activities of a Plethora of Medicinal Plant Extracts and Hydrolates against Human Pathogens and Their Potential to Reverse Antibiotic Resistance. International Journal of Microbiology, 2015; 15:1-15.
- El-Hawary SS, El-Tantawy ME, Rabeh MA and Badr WK. Chemical composition and Biological Activities of Essential oils of A. indica A. Juss. International Journal of Applied Research in Natural Products. 2013; 6(4):33-42.
- Dastan D, Pezhmanmehr M, Askari N, Ebrahimi SN and Hadian J. Essential Oil Composition of the Leaves of Azadirachta indica A. Juss from Iran. Journal of Essential Oil-bearing Plants. 2013; 13(3):357-361.
- Okhale SE, Buba CI, Ogbonna UA and Ettah UAO. Chemical Composition of Essential Oil of Azadirachta indica A. Juss Root from Dikko, Niger State, Nigeria. International Journal of Pharmacognosy. 2018; 5(11):717-717.
- 22. Fritsche KL. The science of fatty acids and inflammation. Advances in nutrition (Bethesda, Md.). 2015; 6(3):293S–301S.
- Lim JH, Gerhart Hines Z, Dominy JE, Lee Y, Kim S, Tabata M, Xiang YK, Puiegserver P. Oleic acid stimulates complete oxidation of fatty acids through protein kinase A-dependent activation of SIRT1-PGCα complex. Journal of Biological Chemistry. 2013; 288:7117-26.
- 24. de Silva PS, Luben R, Shrestha SS, Khaw KT and Hart AR. Dietary arachidonic and oleic acid intake in ulcerative colitis etiology: a prospective cohort study using 7-day food diaries. Eur J Gastroenterol Hepatol. 2014; 26:11-18.
- 25. Ochoa JJ, Pamplona R, Ramirez-Tortosa MC, Granados-Principal S, Perez-Lopez P, Naudi A, Portero-Otin M, Lopez-Frias, Battino M and Quiles JL. Age-related changes in brain mitochondrial DNA deletion and oxidative stress are differentially modulated by dietary fat type and coenzyme Q1. Free Radical Biology and Medicine. 2011; 50:1053-1064.
- Cunnane SC, Schneider JA, Tangney C, Tremblay-Mercier J, Fortier M, Bennett DA and Morris MC. Plasma and brain fatty acid profiles in mild cognitive impairment and Alzheimer's disesase. J Alzheimer's Bis. 2012; 29:691-697.
- 27. Hamazaki K, Hamazaki T and Inadera H. Fatty acid composition in the post-mortem amygdale of patients with schizophrenia, bipolar disorder, and major depressive disorder. Journal Psychiatry Research. 2012; 46:1024-1028.
- 28. Lahey R, Wang X, Carley AN and Lewandowski ED. Dietary fat supply to failing hearts determines dynamic lipid signalling for nuclear receptor activation and oxidation of stored triglyceride. Circulation, 2014; 130:1790-1799.

- 29. Elaigwu ED, Ogo OA, Efiong EE and Oche OG. Effects of ethanolic leaf extracts of neem (Azadirachta indica) on oxidative stability of palm oil. Research Journal Phytochemistry. 2019; 13:1-10.
- 30. Hossain MD, Sarwar MS, Dewan SM, Hossain MS, Shahid-Ud-Daula A and Islam MS. Investigation of total phenolic content and antioxidant activities of Azadirachta indica roots. Avicenna journal of phytomedicine. 2014; 4(2):97–102.
- Ololade ZS, Olawore NO, Olasoji SO and Anosike SO. Chemical Composition and Bactericidal Activities of the Leaf Essential Oil of Eucalyptus Maculata Hook, Natural Product Chemistry and Research. 2017; 5(2):1-4.
- Alao FO, Ololade ZS and Nkeonye CV. Phytochemicals and Antibacterial Potentials of Senna tora Leaf and Seed Extracts on Some Clinically Isolated Bacteria. Journal of Bacteriology and Parasitology. 2018; 9(3):1-4.